

are "sociation" rather than "association" constants and the limitations brought forth by Scott⁸ should be kept in mind when evaluating enthalpies of adduct formation by this procedure. Nevertheless, for systems having enthalpies of adduct formation greater than about 1.5 kcal/mol, these⁸ difficulties should be minimal. The incorrect spectroscopic constants will be obtained only when $\gamma_{DA}/\gamma_D\gamma_A$ is varying or is a constant very much different than unity. To detect the former complication, we strongly advocate treating the data by a Rose-Drago procedure^{4,5,11} and looking at the intersections as a function of concentration.

Acknowledgment. We thank the National Science Foundation for its generous support through U. S. NSF GP 31431X.

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Demonstration of a Direct Hydrogen Transfer between NADH and a Deazaflavin

Sir:

Reductions by NADH have invariably been found to occur by direct transfer of a proton plus two electrons to the substrate molecule.¹ The mechanism of the biochemically important reduction of flavins (7,8-dimethyl-10-alkylisoalloxazines) by NADH is unknown though popular mechanisms reject direct hydride transfer and invoke covalent bond formation between the dihydronicotinamide and the flavin.² The difficulty in determining whether a direct hydride transfer occurs between NADH and flavin is undoubtedly due to the fact that the protons of the ultimate product (1,5-dihydroflavin) are bound to weakly basic nitrogens and are, therefore, exchangeable. Since the 5 nitrogen has been shown, *via* theoretical calculations,³ to be the most electrophilic position of the flavin molecule and, therefore, the most likely recipient of a transferred hydride ion, we have investigated a compound where this nitrogen has been replaced by a carbon. The reaction of 3,10-dimethyl-5-deazaalloxazine (I)⁴ with NADH has been examined in D₂O [80 mg of I suspended in 5 ml of D₂O containing 720 mg of the disodium salt of NADH was stirred for 3 days in the dark (argon atmosphere, 30°); the product (70 mg, 87%) was collected and washed with 2 ml of D₂O]. Except for the deuteron at position 1 the compound obtained was indistinguishable by nmr from that obtained on reduction of I with dithionite in H₂O, the nmr spectrum⁵ establishing conclusively that the reduction product was IsHD(II) of the equation.

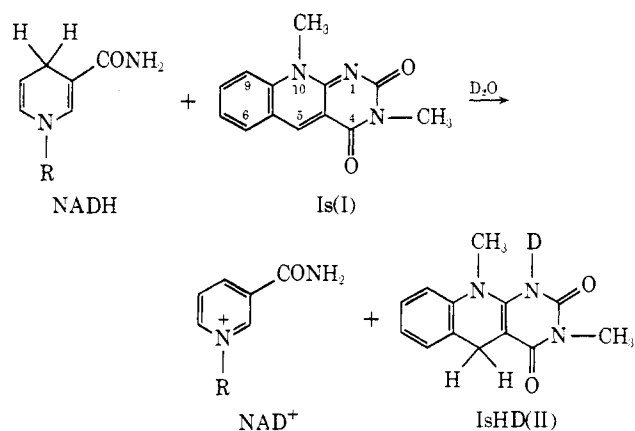
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(2) (a) P. Hemmerich, "Flavins and Flavoproteins," H. Kamin, Ed., University Park Press, Baltimore, Md., 1971, p 103. (b) G. A. Hamilton, *Progr. Bioorg. Chem.*, **1**, 83 (1971).

(3) P.-S. Song, SDN (super delocalizability by nucleophile), FOD, and π_{F} calculations, private communication, 1972.

(4) *Anal.* Calcd for C₁₃H₁₁N₃O₂: C, 64.72; H, 4.60; N, 17.42. Found: C, 64.53; H, 4.55; N, 17.26.

(5) Nmr spectrum of I in CDCl₃: δ 8.95 [1, s, C(5)-H], 8.2-7.2 [4, m, Ar-H], 4.19 [3, s, N(10)-CH₃], 3.47 ppm [3, s, N(3)-CH₃]. Nmr



We have previously presented evidence that reduction of flavins by NADH takes place through a preequilibrium complex in which the dihydronicotinamide does not occupy the area adjacent to the 1, 9, and 10 positions of the flavin.⁶ If the present results are interpretable as evidence for transfer of two electrons from NADH to the flavin and a proton from the 4 position of the NADH to the 5 position of the flavin, then the geometry of the transition state becomes relatively defined. Evidence has been presented for direct hydrogen transfer from NADH to substrate *via* enzyme bound flavin.⁷

In passing it is of interest to note the general similarities of flavins and I. The second-order rate constants for the reactions of NADH and NPrNH with I [$k_{2,\text{NADH}} = 1.89 M^{-1} \text{ min}^{-1}$ at pH 7.62, $k_{2,\text{NPrNH}} = 3.68 \times 10^2 M^{-1} \text{ min}^{-1}$ at pH 7.69 (30°, phosphate buffer containing 5 vol % DMF, $\mu = 0.19$)] are not too dissimilar from the corresponding rate constants obtained with 3,10-dimethylisoalloxazine [$k_{2,\text{NADH}} = 53 M^{-1} \text{ min}^{-1}$, $k_{2,\text{NPrNH}} = 5.25 \times 10^3 M^{-1} \text{ min}^{-1}$].⁶ I reacts with the SO_3^{2-} component of sulfite buffer to provide the 5 adduct,⁸ as previously shown for flavins.⁹ Upon acidification of a sulfite-adduct solution in D₂O with DCl, pure I is generated as proven by the nmr spectrum. IsH₂ regenerates I on reaction with O₂,¹⁰ as do 1,5-dihydroflavins,¹¹ and is oxidized by (CH₃)₂S to yield I. The oxidation of mercaptans by flavins is well established^{6,12,13} and the reduction of a disulfide by IsH₂ is the retrograde of this reaction. As in the case of flavins, I forms nonfluorescent complexes with tryptophan and β -resorcylic acid. The 1:1 complexing constants with tryptophan and β -resorcylic acid, determined by spectrum of II in CDCl₃ (DMSO-*d*₆): δ 7.6-6.8 (7.6-6.8) [4, m, Ar-H], 3.83 (3.63) [2, s, C(5)-H], 3.51 (3.30) [3, s, N(10)-CH₃], 3.38 ppm (3.16) [3, s, N(3)-CH₃]. The proton at position 1 of IsH₂ shows a singlet at δ 4.49 ppm in CDCl₃.

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(7) (a) G. R. Drysdale, *Biochim. Biophys. Acta, Libr.*, **8**, 159 (1966); (b) P. Strittmatter, *ibid.*, **8**, 325 (1966).

(8) 3,10-Dimethyl-5-sulfonate-5-deaza-1,5-dihydroisoalloxazine: λ_{max} 307 nm (ϵ 12,500 $M^{-1} \text{ cm}^{-1}$) at pH 6.78 (10 vol % CH₃CN, $\mu = 0.9$); $K_{\text{eq}} = 3.88 \times 10^2 M^{-1}$ at pH 6.93 (5 vol % DMF, $\mu = 0.19$), 30°; $k_{\text{f}} = 2.09 \times 10^3 M^{-1} \text{ sec}^{-1}$ at pH 6.89 (5 vol % DMF, $\mu = 0.1$), 30°; nmr spectrum in D₂O δ 7.7-6.7 [4, m, Ar-H], 5.13 [1, s, C(5)-H], 3.33 ppm [6, s, N(3,10)-CH₃], the proton at position 1 shows a singlet at δ 5.4 ppm in H₂O.

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(12) I. M. Gascoigne and G. K. Radda, *Biochim. Biophys. Acta*, **131**, 498 (1967).

(13) M. J. Gibian and D. V. Winkelman, *Tetrahedron Lett.*, 3901 (1969).

fluorescence quenching, are 102 and 83 M^{-1} , respectively, for I and 86 and 57 M^{-1} , respectively, for 3,10-dimethylisalloxazine at pH 7.85 (phosphate buffer containing 5 vol % DMF, $\mu = 1.95$).^{13a}

Acknowledgment. This work was supported by a grant from the National Science Foundation.

(13a) NOTE ADDED IN PROOF. Professor P.-S. Song has extended his SDN, FOD, and π_{rr} calculations to I finding the 5 carbon the most electrophilic position. Since for both isalloxazines and 3- and 5-deazaalloxazines the 5 position is the most electrophilic by these criteria, he concludes that the pattern of reactivity of the oxidized flavin should be independent of the atom at the 5 position (N vs. C) (private communication).

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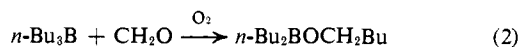
Two Novel Reactions of Monomeric Formaldehyde with Trialkylboranes. A Remarkably Rapid Elimination Diverted by Oxygen to a Free-Radical Chain Addition

Sir:

Monomeric formaldehyde at 0° reacts rapidly with tri-*n*-butylborane to produce 1-butene and methyl di-*n*-butylborinate (eq 1). In the presence of air,



however, the reaction takes another course, producing the one-carbon homologated ester (eq 2). The latter



reaction proceeds through a free-radical chain mechanism involving the intermediates *n*-Bu· and *n*-Bu-CH₂O·. These two reactions appear common to other trialkylboranes.

Trialkylboranes undergo a facile 1,4 addition to α,β -unsaturated carbonyl systems.¹ The reaction involves free-radical intermediates.² No 1,2-addition product has been detected. Indeed, simple organoboranes do not add to the carbonyl group as do other organometallics.³ When an attempt is made to force the addition at 100 to 150°, the reaction follows a different pathway, reductive dealkylation (eq 3).⁴



Such reactions may involve thermal decomposition of the organoborane into R_2BH and olefin,⁵ followed by reaction of the R_2BH with the aldehyde or ketone.

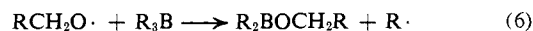
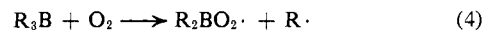
We now wish to report what appears to be the first direct addition of a simple trialkylborane to the carbonyl group. A 100-ml flask equipped with a magnetic stirring bar and septum inlet was charged with 20 ml of mineral oil and 3.0 g of paraformaldehyde (100

mmol). This flask was connected to a dry 100-ml reaction flask, equipped with a magnetic stirring bar and a septum inlet, by a connecting tube and distillation adapter. The vacuum takeoff of the distillation adapter was connected to a mercury bubbler and the system flushed with nitrogen. The reaction flask was cooled to 0° and then charged with 20 ml of tetrahydrofuran (THF) and 10 mmol of tri-*n*-butylborane. The mineral oil was heated to 120°, slowly raised to 140°, to generate monomeric formaldehyde.⁶ After 90 min the borane (gc analysis) had disappeared and a new peak corresponding to methyl di-*n*-butylborinate had appeared. This compound was isolated and identified by comparison with an authentic sample. The reaction mixture was oxidized with 10 ml of 3 *N* sodium hydroxide and 10 ml of 30% hydrogen peroxide. Analysis by gc revealed 20.1 mmol of 1-butanol, 0.3 mmol of 1-pentanol, and 0.2 mmol of tetrahydrofurfuryl alcohol.

The small amount of 1-pentanol indicated that some addition of the tri-*n*-butylborane to the formaldehyde had occurred. The concurrent formation of the tetrahydrofurfuryl alcohol from the solvent suggested that the reaction might involve free-radical intermediates. Many free-radical reactions of organoboranes may be initiated by oxygen.⁷ When the reaction was repeated and air added at 1–2 ml per min through a syringe needle directly above the THF solution, the reaction followed an entirely new path. Oxidation after 90 min produced 20.3 mmol of 1-butanol, 7.8 mmol of 1-pentanol, and 2.6 mmol of tetrahydrofurfuryl alcohol. The use of benzene avoided the formation of the side product.

The presence of iodine⁸ (5 mol %) or copper *N,N*-diethyldithiocarbamate⁹ completely inhibits the formation of the homologated alcohol without affecting the reductive dealkylation. Therefore, the homologation must proceed *via* a free-radical chain reaction while the dealkylation follows a nonradical process.

Oxygen reacts with trialkylboranes to produce alkyl radicals (eq 4). These can add to formaldehyde (eq 5),¹⁰ producing alkoxy radicals capable of displacing alkyl radicals from the organoboranes¹¹ (eq 6).



The mild conditions of the reductive dealkylation suggest that it cannot involve prior dissociation of the organoborane. Instead, the reaction presumably proceeds through a six-member ring transition state similar to that proposed for the rapid dealkylation of organoboranes with *cis*-azobenzene¹² (eq 7). The precise reason why the reaction course is so different for formaldehyde and higher aldehydes is not yet clear.

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